# The Synthesis and Analgesic Activities of Some Spiro[indan-1,3'-pyrrolidine] Derivatives Designed as Rigid Analogs of Profadol

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Abstract  $\Box$  Aromatic hydroxylated derivatives of spiro[indan-1,3'pyrrolidine], designed as conformationally restricted analogs of profadol, were synthesized and pharmacologically evaluated in mice for analgesia and other central nervous system activities. None of the compounds synthesized were as potent as profadol in writhing and hot plate tests, but the 4-hydroxy derivative exhibited codeine-level analgesia in the tests.

Keyphrases □ Profadol—rigid analogs, spiro[indan-1,3'-pyrrolidine] derivatives, synthesis and analgesic activity □ Analgesic activity evaluation of some spiro[indan-1,3'-pyrrolidine] derivatives, synthesis □ Spiro[indan-1,3'-pyrrolidine] derivatives—synthesis and evaluation for analgesic activity

As part of this laboratory's interest in the use of spirocompounds as analgesic receptor probes (1-3), a series of spiro[tetralin-1,3'-pyrrolidine] (I) derivatives, designed as conformationally and rotationally restricted analogs of the analgesic drug, profadol (II), were synthesized recently and their analgesic activities determined. The present report describes the synthesis and analgesic activities of some structurally related spiro[indan-1,3'-pyrrolidine] (III) derivatives in which conformational flexibility is restricted more than in I.

## **EXPERIMENTAL<sup>1</sup>**

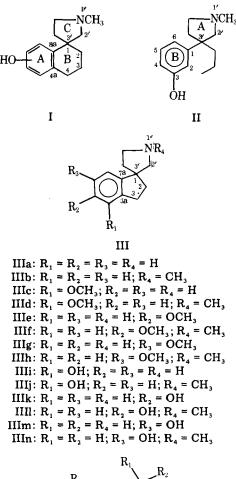
**Chemistry**—Ethyl 4-Methoxy-indanylidenecyanoacetate (IVb)—A mixture of 4-methoxy-1-indanone (16.2 g, 0.1 mole), ethyl cyanoacetate (12.43 g, 0.11 mole), ammonium acetate (15.4 g, 0.2 mole), and acetic acid (48 g, 0.8 mole) in benzene (100 ml) was heated under reflux with a Dean and Stark water trap for 24 hr. The cooled reaction mixture was washed with water ( $3 \times 100$  ml), the washings extracted with benzene ( $2 \times 50$  ml) and the combined benzene liquors dried (magnesium sulfate) and concentrated to give crude product. Recrystallization from ethanol gave an analytical sample (Table I); NMR (dimethyl sulfoxide- $d_6$ ):  $\delta$  1.38 (t, 3H, OCH<sub>2</sub>CH<sub>3</sub>), 2.98 (m, 2H, indan-3 protons), 3.52 (m, indan-2 protons), 3.86 (s, 3H, OCH<sub>3</sub>), 4.32 (q, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 7.00 (1H, m, indan-5 proton), 7.29 (1H, m, indan-6 proton), and 8.26 (1H, m, indan-7 proton) ppm; IR: 2215 (C=N) and 1720 (C=O) cm<sup>-1</sup>.

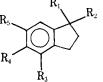
Compounds IVa, IVc, and IVd were prepared similarly from the appropriately substituted 1-indanone<sup>2</sup> (Table I). The IR and NMR spectra were consistent with their assigned structures.

4-Methoxy-1-cyano-1-cyanomethylindane (IVf)—A solution of potassium cyanide (16.2 g, 0.25 mole) in water (50 ml) was added to a stirred solution of IVb (24.5 g, 0.1 mole) in absolute ethanol (200 ml) and the contents stirred at 65° for 16 hr. The mixture was then evaporated to dryness *in vacuo*, the residue suspended in water (200 ml) and extracted with ether (3 × 200 ml). The ether extracts were washed with water (2 × 50 ml), dried (magnesium sulfate), and the solvent evaporated to give a brown oil. This oil was vacuum distilled to give IVf, a colorless oil that crystallized on standing. Recrystallization from ethanol gave an analytical sample (Table I); NMR (deuterochloroform):  $\delta$  2.40 (m, 2H, indan-2 protons), 2.86 (s, 2H, CH<sub>2</sub>CN), 2.97 (m, 2H, indan-3 protons), 3.80 (s, 3H, OCH<sub>3</sub>), 6.83 (m, 1H, indan-5 proton), 7.12 (m, 1H, indan-6 proton), and 7.33 (m, 1H, indan-7 proton) ppm; IR: 2255 and 2240 (both C=N) cm<sup>-1</sup>.

Compounds IVe, IVg, and IVh were prepared similarly from IVa, IVc, and IVd, respectively. IR and NMR spectra were consistent with the assigned structures.

4-Methoxy spiro[indan-1,3'-pyrrolidine-2',5'-dione] (IVj)—A suspension of IVf (21.2 g, 0.1 mole) in acetic acid (50 ml) and 78% (v/v) sulfuric acid (20 ml) was heated in an oil bath at 125° for 1 hr. The acetic acid was removed by *in vacuo* distillation and the resulting mass was suspended in water (50 ml) and extracted with ethyl acetate ( $3 \times 300$  ml). The organic phase was washed with saturated sodium bicarbonate solution ( $2 \times 200$  ml) and water (100 ml) and then dried (magnesium sul-





IVa-IVl (Table I)

<sup>&</sup>lt;sup>1</sup> Melting points were determined on a Reichert hot stage microscope and are uncorrected. IR spectra were determined as Nujol mulls on a Perkin-Elmer 237 spectrophotometer, and NMR spectra were determined on a Perkin-Elmer R12B instrument using tetramethylsilane as the internal standard. <sup>2</sup> 1-Indanone and 5-methoxy-1-indanone were purchased from Aldrich Chemical

<sup>&</sup>lt;sup>2</sup> 1-Indanone and 5-methoxy-1-indanone were purchased from Aldrich Chemical Corp., 4-methoxy-1-indanone was prepared by the method of London and Razdan (4), and 6-methoxy-1-indanone was prepared by the method of House and Hudson (5).

Table I-Physical and Analytical Data for 1,1-Disubstituted Indane Derivatives

Com-									Analysis	
pound	R_1	R <sub>2</sub>	R <sub>3</sub>	R <sub>4</sub>	R <sub>5</sub>	Yield, %	Melting Point	Formula	Calc.	Found
IVa	==C(C	N)CO <sub>2</sub> C <sub>2</sub> H <sub>5</sub>	Н	Н	Н	61.7	103–104°	$C_{14}H_{13}NO_2$	C 74.0 H 5.7 N 6.2	74.2 5.5 6.1
IVb	=C(C	N)CO <sub>2</sub> C <sub>2</sub> H <sub>5</sub>	-OCH <sub>3</sub>	Н	Н	33.1	175–178°	$C_{15}H_{15}NO_3$	C 70.0 H 5.9 N 5.4	70.0 5.9 5.3
IVc	=C(C	N)CO <sub>2</sub> C <sub>2</sub> H <sub>5</sub>	Н	-OCH <sub>3</sub>	Н	50.8	165–167° (subl.)	$C_{15}H_{15}NO_3$	C — H — N —	70.1 6.0 5.3
IVd	=C(C)	N)CO <sub>2</sub> C <sub>2</sub> H <sub>5</sub>	Н	Н	-OCH3	20.4	137–139°	$\mathrm{C}_{15}\mathrm{H}_{15}\mathrm{NO}_3$	C H N	$70.1 \\ 6.0 \\ 5.3$
IVe	CN	CH <sub>2</sub> CN	Н	Н	н	83.5	54–56°	$C_{12}H_{10}N_2$	C 79.1 H 5.5 N 15.4	79.4 5.5 15.4
IVf	CN	CH₂CN	-OCH3	н	Н	84.7	71.5–73°	$C_{13}H_{12}N_2O$	C 73.6 H 5.7 N 13.2	$     \begin{array}{r}       13.4 \\       74.0 \\       5.8 \\       13.2 \\     \end{array} $
IVg	CN	CH <sub>2</sub> CN	Н	OCH <sub>3</sub>	Н	57.7	52–55°	$C_{13}H_{12}N_2O$	C — H — N —	73.4 5.8 12.8
IVh	CN	CH <sub>2</sub> CN	Н	н	OCH <sub>3</sub>	50.8	86–88°	$C_{13}H_{12}N_2O$	К — С — Н — N —	74.1 5.9 13.0
IVi	COI	NHCOCH2-	н	Н	Н	69.7	151–153°	$C_{12}H_{11}NO_2 \\$	C 71.6 H 5.5 N 7.0	$   \begin{array}{r}     13.0 \\     71.7 \\     5.55 \\     6.8 \\   \end{array} $
IVj		NHCOCH2-	OCH <sub>3</sub>	н	Н	61.6	187–189°	$C_{13}H_{13}NO_3$	C 67.5 H 5.7 N 6.1	67.4 5.7 6.0
IVk	-C01	NHCOCH2	н	OCH <sub>3</sub>	Н	66.7	174–176°	$C_{13}H_{13}NO_3$	C H N	67.4 5.4 6.1
IVI	COl	NHCOCH <sub>2</sub>	н	Н	OCH <sub>3</sub>	73.0	154–156°	C <sub>13</sub> H <sub>13</sub> NO <sub>3</sub>	N — C — H — N —	67.6 5.7 5.8

fate). Evaporation of the solvent gave the crude product which was recrystallized from absolute ethanol to give an analytical sample (Table I); NMR (dimethyl sulfoxide- $d_6$ ):  $\delta$  1.87–2.80 (m, 2H, indan-2 protons), 2.89–3.26 (m, 2H, indan-3 protons), 2.03 (s, 2H, pyrrolidine-4' protons), 3.79 (s, 3H, OCH<sub>3</sub>), 6.68 (m, 1H, indan-5 proton), 6.74 (m, 1H, indan-7 proton), 7.21 (m, 1H, indan-6 proton), and 8.50 (broad s, 1H, exchange-able with deuterium oxide, -NH) ppm; IR: 3190, 3075, 1780, 1725, and 1695 cm<sup>-1</sup>.

Compounds IV*i*, IV*k*, and IV*l* were prepared similarly from IV*e*, IV*g*, and IV*h*, respectively; IR and NMR spectra were consistent with the assigned structures.

4-Methoxy spiro[indan-1,3'-pyrrolidine] (IIIc)—Finely divided IVj (23.1 g, 0.1 mole) was added slowly to a stirred suspension of aluminum lithium hydride (11.25 g, 0.3 mole) in dry tetrahydrofuran and the mixture refluxed for 24 hr. The excess aluminum lithium hydride was decomposed by careful addition of water, anhydrous magnesium sulfate was added, and the suspension was filtered. The filtrate was evaporated in vacuo to give a pale yellow oil which was distilled in vacuo to give IIIc; NMR (deuterochloroform):  $\delta$  1.78–2.22 (m, 4H, indan C-2 and C-4' protons), 2.05 (s, 1H, exchangeable with deuterium oxide, NH), 2.69–2.97 (m, 2H, C-3 protons), 2.90 (s, 2H, C-2' protons), 2.07–3.29 (m, 2H, C-5' protons), 3.76 (s, 3H, OCH<sub>3</sub>), 6.66 (m, 1H, C-5 proton), 6.80 (m, 1H, C-7 proton), and 7.17 (m, 1H, C-6 proton) ppm. The base was converted to the fumarate salt which was recrystallized from ethanol-ether to give an analytical sample (Table II).

Compounds IIIa, IIIc, and IIIg were prepared in a similar manner from IV*i*, IV*k*, and IV*l*, respectively and converted to the hydrochloride or fumarate salts (Table II). IR and NMR spectra were consistent with the assigned structures.

4-Methoxy-N-methyl spiro[indan-1,3'-pyrrolidine] (IIId)—A solution of IIIc (2.0 g, 0.01 mole) in formic acid (2.3 g, 0.05 mole) and 40% formaldehyde solution (2 ml, 0.02 mole) was heated with stirring at 95° for 24 hr. The reaction mixture was diluted with 5% hydrochloric acid (5 ml) and extracted with ether (2 × 10 ml). The aqueous phase was made basic with concentrated ammonia solution, and extracted with ether (3 × 20 ml). The combined layers were washed with water (10 ml), dried (magnesium sulfate), and the solvent evaporated. The resulting oil was distilled *in vacuo* to give pure IIId; NMR (deuterochloroform):  $\delta$  1.88–2.30 (m, 4H, C-2 and C-4' protons), 2.35 (s, 3H, N—CH<sub>3</sub>), 2.50–2.98 (m,

4H, C-3 and C-5' protons), 2.64 (s, 2H, C-2' protons), 3.77 (s, 3H, OCH<sub>3</sub>), 6.64 (m, 1H, C-5 proton), 6.90 (m, 1H, C-7 proton), and 7.20 (m, 1H, C-6 proton) ppm. The base was converted to the fumarate salt which was recrystallized from ethanol-ether to give an analytical sample (Table II).

Compounds IIIb, IIIf, and IIIh were prepared in a similar manner from IIIa, IIIe, and IIIg, respectively, and converted to either their hydrochloride or fumarate salts (Table II). Their IR and NMR spectra were consistent with the assigned structures.

4-Hydroxy spiro[indan-1,3'-pyrrolidine]hydrobromide (IIIi)—A solution of IIIc (2.0 g, 0.01 mole) in 48% aqueous hydrobromic acid (25 ml) was refluxed under nitrogen for 1.5 hr. The yellow-brown solution was chilled to give buff crystals of IIIi which were recrystallized from ethanol-ether, giving an analytical sample (Table II); NMR (deuterium oxide):  $\delta$  1.80–2.20 (m, 4H, C-2 and C-4' protons), 2.55–2.90 (m, 2H, C-3 protons), 3.21 (s, 2H, C-2' protons), 3.24–3.68 (m, 2H, C-5' protons), 6.51 (m, 1H, C-5 proton), 6.69 (m, 1H, C-7 proton), and 7.02 (m, 1H, C-6 proton) pm; IR: 3210 (-OH), 2720 (NH) cm<sup>-1</sup>.

Compounds III*j*-III*n* were prepared similarly from the appropriately substituted spiro[indan-1,3'-pyrrolidine]. Their IR and NMR spectra were all consistent with the assigned structures (Table II).

**Pharmacology**—Analgesia was determined by the acetic acid writhing test (6) in groups of six mice. Each group was dosed orally with either vehicle<sup>3</sup> or the compound under test and injected intraperitoneally 30 min later with dilute acetic acid (0.4 ml, 0.25%). Compounds showing analgesia at 50 mg/kg in that test were also tested for analgesia by the hot-plate method (7) (Table II).

Locomotor activity was determined using a battery of photobeam cages  $(30 \times 12 \times 9 \cdot \text{cm})$  traversed by a beam of light. Interruptions of the beam were counted automatically. Groups of 10 mice were dosed and placed singly in the cages. After 45 min the total number of interruptions recorded and the percentage of inhibition or increase in movement relative to controls were determined.

Reversal of reserpine-induced hypothermia was assessed in groups of

<sup>&</sup>lt;sup>3</sup> The testing vehicle used was "Dispersol," ICI Ltd., Pharmaceuticals Division, Macclesfield, UK, which consists of Lissapol NX (Ex ICI Dyestuffs Division, nonylphenol-ethylene oxide concentrate) (0.1%), "Lissapol" C\*\* (sodium salt of sulfated cetyl/oleyl alcohol mixture) (0.1%), "Dispersol" OG\*\* (polyglyceryl ricinoleate) (0.1%), and water (99.7%).

Table II-Physical, Chemical, and Pharmacological Data for Spiro[indan-1,3'-p	•pyrrolidine] Derivatives
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<b>a</b> 1		Melting Point		Analysis	Writhing Test (Mice) ED <sub>50</sub> ,	
Compound	Yield, %		Formula	Calc.	Found	mg/kg po <sup>a</sup>
IIa HCl	79.3	122-125°	C <sub>12</sub> H <sub>15</sub> N·HCl	C 68.7	68.7	>50
				H 7.7	7.6	
	00.4	010 0010		N 6.7	6.6	100(1111100)
IIIb HCl	86.4	219–221°	$C_{13}H_{17}N$ ·HCl	C 69.8 H 8.1	$\begin{array}{c} 69.7 \\ 8.2 \end{array}$	13.9 (11.1–16.3) <sup>b,c</sup>
				N 6.3	6.4	
				C 63.9	64.1	$31.3(23.2-39.6)^d$
IIIc fumarate	98.9	188–195°	C <sub>13</sub> H <sub>17</sub> NO•C <sub>4</sub> H <sub>4</sub> O <sub>4</sub>	H 6.6	6.6	01.0 (20.2 00.0)
-	0010		010-11/0-044-04	N 4.4	4.5	
IIId fumarate	89.2	150–155°	$C_{14}H_{19}NO \cdot C_4H_4O_4$	C 64.9	65.1	NA
				H 6.9	7.1	
				N 4.2	4.2	
IIIe fumarate	89.8	168–175°	$C_{13}H_{17}NO \cdot C_4H_4O_4$	C 63.9	64.0	NA
				H 6.6	6.6	
TTT / f	01.0	140 1500		N 4.4	4.3	N7 A
III/ fumarate	81.9	146–150°	$C_{14}H_{19}NO \cdot C_4H_4O_4$	C 64.9 H 6.9	64.7 6.7	NA
				п 6.9 N 4.2	0.7 3.8	
IIIg fumarate	92.4	124–130°	C <sub>13</sub> H <sub>17</sub> NO·C <sub>4</sub> H <sub>4</sub> O <sub>4</sub>	$\begin{array}{c} 10 & 4.2 \\ C & 63.9 \end{array}$	64.1	NAe
ing fumatate	34.4	124-100	0131117140-0411404	H 6.6	6.6	
				N 4.4	4.25	
IIIh fumarate	93.5	140-147°	C <sub>14</sub> H <sub>19</sub> NO·C <sub>4</sub> H <sub>4</sub> O <sub>4</sub>	C 64.9	64.7	$30.1 (19.1 - 38.2)^d$
			014-019-00 04-04 04	H 6.9	6.7	
				N 4.2	4.0	
III <i>i</i> HBr	75.2	181–187°	C <sub>12</sub> H <sub>15</sub> NO·HBr	C 53.4	53.3	$12.5 \ (8.9-15.1)^{f}$
				H 6.0	6.1	
			~	N 5.2	5.3	
IIIj HBr	75.2	231–237°	C <sub>13</sub> H <sub>17</sub> NO·HBr	C 54.9	55.1	NA
				H 6.4	6.4	
III. IID.	00.0	000 0000		N 4.9	4.7	> 50
IIIk HBr	92.8	222–229°	C <sub>12</sub> H <sub>15</sub> NO•HBr	C 53.4 H 6.0	$53.6 \\ 6.1$	>50
				N 5.2	5.2	
III <i>l</i> HBr	57.3	272–278°	C <sub>13</sub> H <sub>17</sub> NO·HBr	C 54.9	54.9	>50 <sup>g</sup>
	01.0	212-210	01311/110/1101	H 6.4	6.3	200-
				N 4.9	4.9	
IIIm HBr	87.9	263-267°	C <sub>12</sub> H <sub>15</sub> NO•HBr	C 53.4	53.2	NA
			**	H 6.0	5.9	
				N 5.2	5.0	
IIIn HBr	94.3	182–187°	C <sub>13</sub> H <sub>17</sub> NO•HBr	C 54.9	54.8	NA
				H 6.4	6.3	
Morphine				N 4.9	4.9	$1.4 \ (0.7 - 3.6)^h$

<sup>a</sup> Confidence limits in parentheses. <sup>b</sup> Effective at 30 mg/kg in reversing pentylenetetrazol-induced convulsions in mice. <sup>c</sup> Failed to exhibit a normal dose-response curve in the hot-plate test. <sup>d</sup> Not active at 50 mg/kg po in hot-plate test. <sup>e</sup> Exhibited central stimulant activity at 30 mg/kg in mice. <sup>f</sup> ED<sub>50</sub> = 16.8 (11.6–25.3) mg/kg po in hot-plate test. <sup>g</sup> Caused a 90% reduction in motility at 30 mg/kg in mice. <sup>h</sup> ED<sub>50</sub> = 3.8 (2.2–6.5) mg/kg po in hot-plate test. NA Not active at 100 mg/kg.

six mice as described previously (8) and based on the method of Askew (9).

Anticonvulsant activity was measured in groups of six mice subjected to the application of maximal electroshock (20 mamp for 0.22 sec via aural clip electrodes) or administration of the chemical convulsant, pentylenetetrazol (160 mg/kg subcutaneously). In each case, tonic extensor spasm was used as the end-point.

Impairment of motor coordination was assessed by measuring the ability of groups of six mice to balance on a stationary 1.3-cm diameter metal rod. Individual animals were placed on the rod and observed throughout the subsequent 20 sec. The time each mouse remained upon the rod was recorded and converted to a score. If the mouse balanced for the entire 20 sec, a score of 0 was given; if the mouse fell off after T sec, a score of 20 - T was given. Mice that fell off the rod on their first attempt were replaced for three more trials, providing a maximum possible score of 80. The mean score per mouse was determined and compared by standard statistical procedures with corresponding values obtained for groups treated with vehicle alone.

## RESULTS

**Chemistry**—The syntheses of compounds IVa-IVl and IIIa-IIIh were carried out using procedures similar to those described previously for the preparation of the spiro[tetralin-1,3'-pyrrolidine] derivatives (3), but utilizing the appropriately substituted 1-indanone in place of the corresponding 1-tetralone. The appropriate 1-tetralone was condensed with ethylcyanoacetate to give the 1-indanylidenecyanoacetate derivative (IVa-IVd), which was reacted with potassium cyanide to give the corresponding 1-cyano-1-(cyanomethyl)indane (IVe-IVh). Treatment of compounds IVe-IVh with an acetic acid-sulfuric acid mixture gave the

corresponding spiro[indan-1,3'-pyrrolidine-2',5'-dione] derivatives (IVi–IVI). These compounds were then reduced with aluminum lithium hydride in tetrahydrofuran to give the appropriately substituted spiro[indan-1,3'-pyrrolidine] (III). O-Demethylation of compounds IIIc–IIIh was carried out under nitrogen in refluxing 48% hydrobromic acid containing 1% phosphoric acid. N-Methylation of compounds IIIa, IIIc, IIIe, and IIIg was accomplished in formic acid–formaldehyde solution (Tables I and II).

**Pharmacology**—Compounds III*b* and III*i* were the most active compounds in the series, exhibiting codeine-level analgesia, whereas compound III*j* showed no analgesic activity. None of the other hydroxylated derivative (III*k*-III*n*) showed any significant analgesic activity, although compound III*l* caused marked reduction in motility in mice at a 30-mg/kg dose, whereas compound III*g* exhibited central stimulant properties at 30 mg/kg in mice. Compound III*b* protected mice against pentylenetetrazol induced convulsions at 30 mg/kg. Compounds III*a*-III*n* were inactive in the maximal electroshock test, in reserpine antagonism tests, and did not produce any impairment of coordination in the stationary rod test.

### DISCUSSION

An important feature of profadol (II) structure is the rotational freedom present in the molecule which allows a number of rotameric forms as the aromatic ring is rotated about the C1–C3' bond. Thus, the weak analgesic activity exhibited by compounds of structure I possibly results from these compounds adopting conformations that are noncomplimentary with the receptor. Spiro[indan-1,3'-pyrrolidine] (III) derivatives may be regarded as profadol analogs, which are more conformationally rigid than the spiro system (I), since aromatic ring rotation is severely restricted in III as a result of the reduction of ring B to a five-membered ring. Thus, compounds IIIj, IIIl, and IIIn represent interesting rigid analogs of II. Furthermore, molecular model examination of IIIj and morphine shows the phenolic and tertiary amino groups, and the aromatic ring in both compounds to be almost stereosuperimposable. In addition, it was of interest to find a patent describing some 3-hydroxyspiro[indan-1,3'-pyrrolidine] derivatives as orally active analgesics (10). Profadol is thought to interact with the analgesic receptor in the halfchair conformation (11).

Although the spiro systems I and III represent structural analogs in which the profadol molecule is fixed in a conformation related to the morphine molecule, these compounds exhibit only weak or insignificant analgesic properties. Thus, other structural features appear to be important in profadol-receptor interaction. The inability of these spiro analogs to enhance or maintain the analgesic properties of profadol may be related to a decreased fit of these compounds at the receptor due to the rigid orientation of the B ring in these compounds, which represents a fixed, and perhaps undesirable, conformation of the 3-propyl group in II.

#### REFERENCES

(1) P. A. Crooks and H. E. Rosenberg, J. Med. Chem., 21, 585 (1978).

(2) P. A. Crooks and H. E. Rosenberg, J. Chem. Soc., Perkin I, 1979, 2719.

- (3) P. A. Crooks and R. Szyndler, J. Med. Chem., 23, 679 (1980).
- (4) J. D. London and R. K. Razdan, J. Chem. Soc., 1954, 4299.
- (5) H. O. House and C. B. Hudson, J. Org. Chem., 35, 647 (1970).

(6) C. Vander Wende and S. Margolin, Fed. Proc., Fed. Am. Soc. Exp. Biol., 15, 494 (1956).

(7) J. A. J. Jannsen and A. H. Ingeneau, J. Pharm. Pharmacol., 9, 381 (1957).

(8) D. T. Greenwood and A. R. Sommerville, Psychopharmacologia, 24, 231 (1972).

(9) B. M. Askew, Life Sci., 2, 725 (1963).

(10) J. M. Bastian, K. Hasspacker, and M. Strasser, Ger. Offen., 2,241,027 (1973); through Chem. Abstr., 78, 159422W (1974).

(11) A. S. Horn and J. R. Rodgers, Nature (London), 260, 795 (1976).

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# Quantitation of Hydroxyprogesterone Caproate, Medroxyprogesterone Acetate, and Progesterone by Reversed-Phase High-Pressure Liquid Chromatography

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Abstract A high-pressure liquid chromatographic method for the quantitation of hydroxyprogesterone caproate, medroxyprogesterone acetate, and progesterone in pharmaceutical dosage forms was developed. The method gave accurate, precise, and reproducible results. The excipients present in the dosage forms did not interfere with the assay procedure except benzyl benzoate in progesterone injection. The percent relative standard deviations based on six injections were 1.6, 2.5, and 2.7% for hydroxyprogesterone caproate, medroxyprogesterone acetate, and progesterone, respectively. The stability of progesterone in ethanolpropylene glycol-water (10:50:40) was studied. The loss in potency of progesterone, even after 487 days of storage at 50°, was <10%.

**Keyphrases** Hydroxyprogesterone caproate—quantitation by reversed-phase high-pressure liquid chromatography 
Medroxyprogesterone acetate-quantitation by reversed-phase high-pressure liquid chromatography **D** Progesterone—quantitation by reversed-phase high-pressure liquid chromatography 
High-pressure liquid chromatography-quantitation of hydroxyprogesterone caproate, medroxyprogesterone acetate, and progesterone

The pharmaceutical dosage forms of hydroxyprogesterone caproate (I), medroxyprogesterone acetate (II), and progesterone (III) are used extensively. The USP method (1) for the quantitative determination of I in dosage forms is based on a reaction with isoniazid. The color produced is measured spectrophotometrically.

For dosage forms of II, the USP method (2) requires normal phase high-pressure liquid chromatography (HPLC) with a porous silica column. The quantitation of III requires reversed-phase chromatography (3). The 1-m column for this procedure has to be packed with octade-

cylsilane chemically bonded to silica gel. The USP method (3) appears to be a modification of a procedure recommended previously (4).

This investigation attempted to develop a reversedphase HPLC method (with prepacked column) suitable for the quantitation of I, II, and III in pharmaceutical dosage forms. The stability of III in some aqueous systems was also determined.

#### EXPERIMENTAL

Reagents and Chemicals-All reagents and chemicals were ACS, USP, or NF grade and were used as received. 17-Hydroxyprogesterone<sup>1</sup> hydroxyprogesterone caproate<sup>1</sup>, medroxyprogesterone acetate<sup>2</sup>, and progesterone<sup>3</sup> were used without further purification.

Apparatus—The chromatograph<sup>4</sup> was connected to a multiplewavelength detector<sup>5</sup>, a recorder<sup>6</sup>, and an integrator<sup>7</sup>. All pH values were determined using a pH meter<sup>8</sup>.

Column—The column<sup>9</sup> (30 cm  $\times$  4-mm i.d.) was of a semipolar material consisting of a monomolecular layer of cyanopropylsilane permanently bonded by silicone-carbon bonds.

Chromatographic Conditions-The mobile phase contained 30%

E. R. Squibb & Sons, Princeton, N.J.
 <sup>2</sup> The Upjohn Co., Kalamazoo Mich.
 <sup>3</sup> Aldrich Chemical Co., Milwaukee, Wis.
 <sup>4</sup> Model ALC 202 equipped with a U6K universal injector, Waters Associates, Milford, Mass.

 <sup>&</sup>lt;sup>6</sup> Spectroflow monitor SF770, Schoeffel Instruments Corp., Westwood, N.J.
 <sup>6</sup> Omniscribe 5213-12, Houston Instruments, Austin, Tex.
 <sup>7</sup> Autolab minigrator, Spectra-Physics, Santa Clara, Calif.
 <sup>8</sup> Model 4500 digital pH meter, Beckman Instruments, Irvine, Calif.
 <sup>9</sup> µBondapak/CN, Waters Associates, Milford, Mass.